Small Angle X-ray Scattering (SAXS) and Biological Applications

Zehra Sayers
Sabanci University, Istanbul, Turkey
SESAME, Chair, Scientific Advisory Committee
Overview

- Protein folding and structure.
- Principles of Small Angle X-ray Scattering (SAXS).
- SAXS measurements on biological samples.
- *Ab initio* modeling
  Heterotrimeric G-proteins of A. Thaliana
  Metallothioneins from wheat.
Levels of Protein Folding

Primary structure: Linear amino acid sequence; directional.

N-Terminal → MSVELKERHAVA.........., KIWAFGGHRRVI → C-Terminal

Secondary structure: Regions with defined fold;
alpha helices and beta sheets

Tertiary structure: Fully folded 3D molecular structure of a single chain.

Quaternary structure: 3D structure of a multi-chain molecule.
Folding process: transition from the high-energy unfolded state to low energy folded state.

A number of metastable intermediate states are sampled before folded state is reached. In solution there may be a dynamic equilibrium of different conformations.
Protein crowding in cells:

Maximum concentration 300-500mg/ml.
Macromolecular Structure Determination

X-ray Crystallography

Snapshots of the 3D structure at atomic resolution. Static measurements*

NMR

Determination of 3D solution structure at high resolution.

SAXS
Modeling of molecular shape envelope at low resolution. Determination of structural parameters e.g. Radius of gyration ($R_g$), molecular mass (MM) etc.

Dynamic measurements to detect changes in structure upon a perturbation.
Interactions of X-Rays with Matter

- Coherent scattering; Structural information at the atomic/molecular level.
- Absorption, fluorescence, near edge measurements: Material characterization, local structure, coordination.
- Transmission/phase contrast: Lower resolution imaging.
Reciprocity law of scattering; inverse relationship between particle size and scattering angle.

Dimensions of biological macromolecules ($D_{\text{max}}$) >> wavelength of X-rays ($\lambda$).

Scattering takes place at low angles.

Inhomogeneities in electron density in a solution of macromolecules in buffer $\Rightarrow$ small angle X-ray scattering (coherent scattering).
Scattering Curves and Particle Size

Scattering angle $2\theta$.
Path difference $1\lambda$.

Destructive interference;
No scattering

Scattering angle $< 2\theta$
Scattering.

Scattering angle $= 0$
Maximum scattering. (Curve 1)

Effect of particle size:

Large particles
path difference $1\lambda$ occurs at smaller angles (Curve 2).

Glatter, O. And Kratky, O. (1982)
Interference and Coherent Scattering

In coherent scattering the path length difference between waves scattered at different electrons is fixed and amplitudes are added.

Path difference = \( \mathbf{r} \cdot \mathbf{S} - \mathbf{r} \cdot \mathbf{S}_0 \)

The total amplitude from two centers (one at the origin and one at \( r \)) is:

\[
F(s) = \sum_{i=1}^{2} f_e \exp(2\pi i s \mathbf{r}_i) = f_e + f_e \exp(2\pi i s \cdot \mathbf{r}_2)
\]
Scattering from Crystals vs from Solutions

\[ F(s) = \sum_{i=1}^{N} f_i(s) \exp(2\pi i s \cdot r_i) \]

"Structure factor"

Fourier transform of the distribution of the spherical atoms.

In SAXS \( F(s) \) refers to structure of the solution; solvent + homogeneous distribution of proteins

The intensity:

\[ I(s) = \sum_{i=1}^{N} \sum_{j=1}^{N} f_i(s) f_j(s) \exp(2\pi i s \cdot (r_i - r_j)) \]

Crystal structure

In solution particles are randomly oriented

\[ < \exp \left( 2\pi i s \cdot (r_i - r_j) \right) > = \frac{\sin(2\pi sr_{ij})}{2\pi sr_{ij}} \]

\[ I(s) = \sum_{i=1}^{N} \sum_{j=1}^{N} f_i(s) f_j(s) \frac{\sin(2\pi sr_{ij})}{2\pi sr_{ij}} \]

Debye (1915)
Contribution of $r_{ij}$ to the Scattering Pattern

$$I(s) = \sum_{i=1}^{N} \sum_{j=1}^{N} f_i(s)f_j(s) \frac{\sin(2\pi sr_{ij})}{2\pi sr_{ij}}$$

$d = r_{ij}$

Short distances; low frequencies dominate.

Large distances; high frequencies dominate higher angles.
I(s) and Distance Distribution Function $P(r)$

$$I(s) = 4\pi \int_0^{D_{\text{max}}} p(r) \frac{\sin(2\pi sr)}{2\pi sr} \, dr$$

For a homogeneous particle $p(r)$: the histogram of distances between pairs of points within the particle.

Scattering intensity and $p(r)$ are related by a Henkel transformation.
Scattering intensity is the absolute square of the resultant amplitude.

In contrast with a diffraction pattern it is a continuous function.

Scattering Intensity

BSA
4.3 mg/ml
Hepes buffer pH 7.0
1 mM DTT
I(s) and Structural Parameters

I(s) is dependent on the molecular shape and size.

\[ I(s) = I(0) \exp\left(-s^2 R_g^2/3\right) \]

Guinier approximation:

- \( R_g \) is the radius of gyration for the particle.
- Plot of \( \ln I(s) \) vs \( s^2 \).

I(0) proportional to the molecular mass (MM) of protein.

Determine with respect to protein with known MM.

- BSA
  - 4.3 mg/ml
  - Hepes buffer pH 7.0
  - 1 mM DTT
I(s) and Structural Parameters

Porod Volume

$I(s)s^4$ vs $s$; particle volume

supplementary information on molecular mass (MM).

$\text{BSA}$

4.3 mg/ml

Hepes buffer pH 7.0

1 mM DTT

$\text{MM is estimated as } \frac{1}{2} \text{ Porod volume.}$
I(s) and Structural Parameters

Kratky Plot

$I(s)s^2$ vs $s$; information on shape e.g. globular or extended folded or natively unfolded, flexible or rigid structure.

Unfolded proteins would display a monotonously increasing Kratky plot.

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BSA
4.3 mg/ml
Hepes buffer pH 7.0
1 mM DTT
Contrast

Particle: \[ F_p(s) = \int_{V} \rho_p(r) \exp(2\pi is \cdot r) dr \]

Solvent: \[ F_b(s) = \rho_b \int_{V} \exp(2\pi is \cdot r) dr \]

Only fluctuations in electron density contribute to the scattering:

\[ I_{obs}(s) = I_p(s) - I_b(s) \]

“Contrast matching”
SAXS Beamline Basics

**Conditions:**

\[ d \ll D \quad \Delta \lambda / \lambda \leq 0.1 \]

\[ \sin 2\theta = 2\theta \quad \cos 2\theta = 1 \]

\( d \): distance on the detector, \( D \): Sample-Detector distance  
WAXS: Wide angle X-ray Scattering

\( X33 \) Small angle X-ray scattering instrument of EMBL in HASYLAB.
**X33 Beamline @ EMBL Hamburg Outstation**

**DORIS III/DESY 4.4 GeV, 120 mA.**

Automatic sample changer, sample can be kept under anaerobic conditions during measurements.

Marr/Pilatus detector, basic data reduction coupled to data collection.
Basics of SAXS Data Reduction

\[
I(S) = \frac{1}{c} \left[ \frac{I_s(S)}{I_{s,0}} - \frac{I_b(S)}{I_{b,0}} \right] \frac{1}{D(S)}
\]

$I(s)$ scattered intensity,
$I_s(s)$ scattering from the sample,
$I_b(s)$ scattering from the buffer,
$c$ concentration of the sample,
$I_{s,0}$ and $I_{b,0}$ through beam for sample and buffer respectively,
$D(s)$ detector response.

Data quality:
- aggregation
- polydispersity
- improper background subtraction
- concentration correction
Analyzing SAXS Data

Log I(s)

Resolution, nm

Size

Shape

Fold

Atomic structure

© Dmitri Svergun
Ab initio Shape Determination

Low Resolution 3D Models

Envelope function

Chacón, P. et al. (1998) Biophys. J. 74, 2760

Bead models


Dummy residues model

These methods all minimize Discrepancy[Data] + Penalty[Additional info]
Oligomeric Forms and Missing Domains

Validation of modeling using simulated data from glutamyl-tRNA synthetase (GTS) complexed with tRNA (1g59)

SASREF for reconstructing oligomeric structures.
BUNCH for reconstructing with missing domains.

SAXS Based Information

Shape determination and low resolution structural analysis for bio-molecules that do not crystallize.

Combination with PX data from homologs to obtain structural information.

Combination with PX data to obtain structural information about missing domains.

Structural analyses of large complexes.

Investigation of intermediates during assembly.

Investigation of shape changes in response to perturbations.
SAXS Measurements with Plant Proteins

Abiotic stress responses in plants

Durum wheat metallothionein dMT
  Small molecular weight ~7 kDa.
  Does not crystallize.

Heterotrimeric G-protein subunits from *Arabidopsis thaliana*
  Complex of three subunits molecular weight ~100 kD.
  Modified forms from mammalian cells crystallize.
  Large structural changes upon interactions. Studies in solution
Durum Wheat Metallothionein (dMT)

-Metallothionein (mt) gene identified in durum wheat tolerant to cadmium. (Metal stress)

-Recombinant protein is produced in bacteria as fusion with GST (GSTdMT) for purification with and without GST, characterization and mutations.

At 25 μM Cd.

dMT Amino acid sequence: 7 kDa

MSCNCGCSCGSDCKCGKMYPDLTEQGSAAQVAAVVLGVAPENKAGQFEVAAGQSGEGCSCGDNCCKCNPCNC

N-terminal  Hinge region  C-terminal
β-domain  +  α-domain

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 Shoot and root growth of the durum wheat cultivar Balcalı-85 with increasing Cd application.

Northern blot analysis of Cd dose dependent expression of dmt gene in roots of *T. durum*.

dMT synthesis is induced during Cd-response

Bilecen et al 2005
Dede et al 2006
Yesilirmak and Sayers 2009
Aydin et al 201 (in preparation)
dMT Purification and Characterization

**Size Exclusion Chromatography**

- **Holo-dMT:** 10-16 kDa
- **Apo-dMT:** 8-9 kDa

**SDS- and Native-PAGE Analysis**

**DLS Measurements**
dMT Purification and Characterization

UV Absorbance Measurements

Circular Dichroism Spectropolarimetry Measurements

Cadmium Content (ICP-OES)

$5.3 \pm 0.5 \text{ Cd ions/dMT}$
Pushing SAXS to its Limits: Scattering from dMT

Scattered Intensity

P(R) Functions

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**Scattered Intensity**

**P(R) Functions**

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**Structural Parameters**

<table>
<thead>
<tr>
<th></th>
<th>Conc (µM)</th>
<th>( R_g ) (nm)</th>
<th>( D_{\text{max}} ) (nm)</th>
<th>( I(0) )</th>
<th>( \text{MM}_{\text{ex}}^{\text{p}} ) (kDa)</th>
<th>( \text{MM}_{h}^{\text{t}} ) (kDa)</th>
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<tbody>
<tr>
<td>Holo-dMT</td>
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<td>7</td>
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<td>11</td>
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<tr>
<td>Apo-dMT pH 8.3</td>
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<td>2.66</td>
<td>8.9</td>
<td>6.8</td>
<td>8.1</td>
<td>7.89</td>
</tr>
</tbody>
</table>

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**X33 SAXS Beamline @ EMBL Hamburg Outstation, HASYLAB**

**Atsas Software, EMBL Hamburg**
Durum Wheat Metallothionein (dMT)

Homology modeling

Known homologs for metal-binding domains, *Ab initio* for the hinge region

Elongated two domain structure with a folded hinge region
Ab initio Models of Apo- and Holo-dMT

Holo-dMT is stable as staggered dimer in the presence of Cd$^{2+}$. Oligomeric forms appear to be responsible for more efficient removal of toxic metals.
SUPPORT

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