## Small Angle X-ray Scattering (SAXS) and Biological Applications

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### Overview

-Protein folding and structure.

- -Principles of Small Angle X-ray Scattering (SAXS).
- -SAXS measurements on biological samples.
- -*Ab initio* modeling Heterotrimeric G-proteins of A. Thaliana Metallothioneins from wheat.

## Levels of Protein Folding

Primary structure: Linear amino acid sequence; directional.

 $N-Terminal \rightarrow MSVELKERHAVA....KIWAFGGHRRVI \rightarrow C-Terminal$ 

Secondary structure: Regions with defined fold; alpha helices and beta sheets

Tertiary structure: Fully folded 3D molecular structure of a single chain.

Quaternary structure: 3D structure of a multi-chain molecule.

# **Protein Folding**





Folding process: transition from the high-energy unfolded state to low energy folded state.

A number of metastable intermediate states are sampled before folded state is reached. In solution there may be a dynamic equilibrium of different conformations.

## **Protein Structure in Solution**



### Protein crowding in cells: Maximum concentration 300-500mg/ml.

## Macromolecular Structure Determination

X-ray Crystallography

Snapshots of the 3D structure at atomic resolution. Static measurements\*

### NMR

Determination of 3D solution structure at high resolution.

### SAXS

Modeling of molecular shape envelope at low resolution. Determination of structural parameters e.g. Radius of gyration ( $R_q$ ), molecular mass (MM) etc.

Dynamic measurements to detect changes in structure upon a perturbation.

### Interactions of X-Rays with Matter



-Coherent scattering; Structural information at the atomic/molecular level.

-Absorption, fluorescence, near edge measurements: Material characterization, local structure, coordination.

-Transmission/phase contrast: Lower resolution imaging.

## Small Angle X-ray Scattering (SAXS)

#### Macromolecules



### Reciprocity law of scattering;

inverse relationship between paticle size and scattering angle.

Dimensions of biological macromolecules  $(D_{max})$  >> wavelength of X-rays ( $\lambda$ ).

 $^{
m >}$  Scattering takes place at low angles.

Inhomogeneities in electron density in a solution macromolecules in buffer  $\Rightarrow$  small angle X-ray scattering (coherent scattering).

### Scattering Curves and Particle Size





I2 $2\theta$  Scattering angle  $2\theta$ . Path difference  $1\lambda$ .

Destructive interference; No scattering

Scattering angle  $< 2\theta$ Scattering..

Scattering angle = 0 Maximum scattering. (Curve 1)

Effect of particle size:

Large particles path difference  $1\lambda$  occurs at smaller angles (Curve 2).



### Scattering from Crystals vs from Solutions

$$\mathbf{F}(\mathbf{s}) = \sum_{i=1}^{N} \mathbf{f}_{i}(\mathbf{s}) \exp(2\pi \, \mathbf{i} \, \mathbf{s} \cdot \mathbf{r}_{i})$$

"Structure factor"

Debye (1915)

Fourier transform of the distribution of the spherical atoms.

In SAXS F(s) refers to structure of the solution; solvent + homogeneous distribution of proteins

The intensity:  

$$I(\mathbf{s}) = \sum_{i=1}^{N} \sum_{j=1}^{N} f_i(s) f_j(s) \exp(2\pi i \mathbf{s} \cdot (\mathbf{r}_i - \mathbf{r}_j)) \longrightarrow Crystal structure$$

In solution particles are randomly oriented

$$\langle \exp (2\pi i \mathbf{s} \cdot (\mathbf{r}_i - \mathbf{r}_j) \rangle = \frac{\sin (2\pi s r_{ij})}{2\pi s r_{ij}}$$

$$I(s) = \sum_{i=1}^{N} \sum_{j=1}^{N} f_i(s) f_j(s) \frac{\sin(2\pi s r_{ij})}{2\pi s r_{ij}}$$

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For a homogeneous particle p(r): the histogram of distances between pairs of points within the particle.

Scattering intensity and p(r) are related by a Henkel transformation.

### Scattering Intensity



-Scattering intensity is the absolute square of the resultant amplitude.

-In contrast with a diffraction pattern it is a continuous function.

### I(s) and Structural Parameters

I(s) is dependent on the molecular shape and size.



I(0) proportional to the molecular mass (MM) of prote Determine with respect to protein with known MM.

Guinier approximation:  $I(s) = I(0)exp(-s^2R_g^2/3)$  $sR_g \lesssim 1.3$ 

Rg is the radius of gyration for the particle. Plot of LnI(s) vs s<sup>2</sup>.

# I(s) and Structural Parameters

Porod Volume

### I(s)s<sup>4</sup> vs s; particle volume supplementary information on molecular mass (MM).



MM is estimated as  $\frac{1}{2}$  Porod volume.

## I(s) and Structural Parameters

Kratky Plot

 $I(s)s^2 vs s$ ; information on shape e.g. globular or extended

folded or natively unfolded, flexible or rigid structure.



Unfolded proteins would display a monotonously increasing Kratky plot.

# Contrast

Particle: 
$$F_p(\mathbf{s}) = \int_V \rho_p(\mathbf{r}) \exp(2\pi i \mathbf{s} \cdot \mathbf{r}) d\mathbf{r}$$

Solvent: 
$$F_b(\mathbf{s}) = \rho_b \int_V \exp(2\pi i \mathbf{s} \cdot \mathbf{r}) d\mathbf{r}$$

Only fluctuations in electron density contribute to the scattering:

$$\mathbf{I}_{obs}(\mathbf{s}) = \mathbf{I}_{p}(\mathbf{s}) - \mathbf{I}_{b}(\mathbf{s})$$

### "Contrast matching"

### **SAXS** Beamline Basics WAXS Data acquisition detector mirror source monochromator 20 slits SAXS sample slits detector $s=2\sin\theta/\lambda$ Scattering vector: $\mathbf{s} = \mathbf{S} \cdot \mathbf{S}_0$ Conditions: d<<D $\Delta\lambda/\lambda \leq 0.1$ $sin2\theta = 2\theta$ $\cos 2\theta = 1$ d:distance on the detector, D: Sample-Detector distance WAXS: Wide angle X-ray Scattering X33 Small angle X-ray scattering instrument of EMBL in HASYLAB.

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# X33 Beamline @ EMBL Hamburg Outstation

DORIS III/DESY 4.4 GeV, 120 mA.



Marr/Pilatus detector, basic data reduction coupled to data collection.

Automatic sample changer, sample can be kept under anaerobic conditions during measurements.



## **Basics of SAXS Data Reduction**



$$I(s) = \frac{1}{c} \left[ \frac{I_s(s)}{I_{s,0}} - \frac{I_b(s)}{I_{b,0}} \right] \frac{1}{D(s)}$$

I(s) scattered intensity, I<sub>s</sub>(s) scattering from the sample, I<sub>b</sub>(s) scattering from the buffer, C concentration of the sample, I<sub>s,0</sub> and I<sub>b,0</sub> through beam for sample and buffer respectively, D(s) detector response.

Data quality:

aggregation polydispersity improper background subtraction concentration correction

## Analyzing SAXS Data



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These methods all minimize Discrepancy[Data] + Penalty[Additional info]

### **Oligomeric Forms and Missing Domains**

Validation of modeling using simulated data from glutamil-tRNA synthetase (GTS) complexed with tRNA (1g59)





SASREF for reconstructing oligomeric structures. BUNCH for reconstructing with missing domains.

Petoukhov, M. V. And Svergun, D. (2005)

# SAXS Based Information

Shape determination and low resolution structural analysis for bio-molecules that do not crystallize.

Combination with PX data from homologs to obtain structral information.

Combination with PX data to obtain structural information about missing domains.

Structural analyses of large complexes.

Investigation of intermediates during assembly.

Investigation of shape changes in response to perturbations.

### SAXS Measurements with Plant Proteins

Abiotic stress responses in plants

Durum wheat metallothionein dMT

Small molecular weight ~7 kDa. Does not crystallize.

Heterotrimeric G-protein subunits from Arabidopsis thaliana Complex of three subunits molecular weight ~100 kD. Modified forms from mammalian cells crystallize. Large structural changes upon interactions. Studies in solution

# Durum Wheat Metallothionein (dMT)



At 25 μM Cd.

-Metallothionein (*mt*) gene identified in durum wheat tolerant to cadmium. (Metal stress)

-Recombinant protein is produced in bacteria as fusion with GST (GSTdMT) for purification with and without GST, characterization and mutations.

### dMT Amino acid sequence: 7 kDa

N-terminal β-domain Hinge region

 $\begin{array}{c} \textbf{C-terminal} \\ \alpha \textbf{-domain} \end{array}$ 

# dMT Expression Under Cadmium Stress





Northern blot analysis of Cd dose dependent expression of d*mt gene* in roots of *T. durum*.

Shoot and root growth of the durum wheat cultivar Balcalı-85 with increasing Cd application.

Bilecen et al 2005 Dede et al 2006 Yesilirmak and Sayers 2009 Aydin et al 201 (in preparation)

### dMT synthesis is induced during Cd-response

### dMT Purification and Characterization

#### Size Exclusion Chromatography



#### SDS- and Native-PAGE Analysis





#### Holo-dMT: 10-16 kDa Apo-dMT: 8-9 kDa





### dMT Purification and Characterization

UV Absorbance Measurements

Circular Dichroism Spectropolarimetry Measurements





### Structural Parameters

	Conc (µM)	R <sub>g</sub> (nm)	D <sub>max</sub> (nm)	I(0)	MM <sub>ex</sub> (kDa)	MM <sub>t</sub> (kDa)
Holo-dMT	192	2.10	7	14.25	17	16.9
Apo-dMT pH 2.5	174	2.98	11	7.6	9.1	7.89
Apo-dMT pH 8.3	174	2.66	8.9	6.8	8.1	7.89

### X33 SAXS Beamline @ EMBL Hamburg Outstation, HASYLAB Atsas Software, EMBL Hamburg

# Durum Wheat Metallothionein (dMT)

Homology modeling

Known homologs for metal-binding domains, *Ab initio* for the hinge region



Elongated two domain structure with a folded hinge region





Holo-dMT is stable as staggered dimer in the presence of Cd<sup>2+</sup>. Oligomeric forms are appear to be responsible for more efficient removal of toxic metals.

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